

Ecology

Community structure of Ichneumonidae (Hymenoptera) in a mangrove area in the coastal zone of Tamaulipas, Mexico

Estructura de la comunidad de Ichneumonidae (Hymenoptera) en un área de manglar en la zona costera de Tamaulipas, México

Blas Antonio Pérez-Urbina ^a, Juana María Coronado-Blanco ^b, Enrique Ruíz-Cancino ^b, Crystian Sadiel Venegas-Barrera ^a, Alfonso Correa-Sandoval ^a, Jorge Víctor Horta-Vega ^{a,*}

^a Instituto Tecnológico de Ciudad Victoria, Blvd. Emilio Portes Gil 1301 Pte., 87010 Cd. Victoria, Tamaulipas, Mexico

^b Facultad de Ingeniería y Ciencias, Universidad Autónoma de Tamaulipas, Centro Universitario Adolfo López Mateos, 87149 Cd. Victoria, Tamaulipas, México

*Corresponding author: jhortavega@yahoo.com.mx (J.V. Horta-Vega)

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Abstract

The structure of Ichneumonidae communities in a mangrove area and 2 nearby locations with different vegetation types are described. The study area is located within the limits of the Nearctic and Neotropical regions in the southern part of the state of Tamaulipas, Mexico. Samples were collected with a Malaise trap at each site over a one-year period. We estimated the potential species richness with the Clench model, the Simpson and Shannon-Wiener diversity indexes and the possible differences among communities at different sites using Permanova multivariate analysis. The relative degrees of influence of space, time, and biogeographical distribution on the community structures of ichneumonid wasps were determined using multiple correspondence analyses. Our results showed that the mangrove area had the highest potential species richness, and that there were significant differences among the 3 ichneumonid communities. The community structures depend primarily on the biogeographical distribution of species and secondarily on the month of collection, whereas the type of vegetation contributed with lower variation. Three species obtained in this study are new records for Mexico, and 5 for Tamaulipas.

Keywords: Correspondence analysis; Parasitic wasps; Neotropical; Mangrove; Biogeographical provinces

Resumen

Se describe la estructura de las comunidades de Ichneumonidae en un área de manglar y 2 localidades cercanas con diferentes tipos de vegetación. El área de estudio está ubicada en los límites de las regiones Neártica y Neotropical en sur del estado de Tamaulipas, México. Las muestras se recolectaron con una trampa Malaise en cada sitio durante 1 año. La riqueza potencial de especies se determinó con el modelo de Clench, los índices de diversidad de Simpson y Shannon-Wiener, así como las posibles diferencias entre las comunidades con el análisis multivariado de Permanova. Los grados relativos de influencia de las variables espaciales, de tiempo y de distribución biogeográfica en las estructuras comunitarias se determinaron mediante análisis de correspondencia múltiple. Los datos mostraron que el área de manglar tiene la mayor riqueza potencial de especies y que hubo diferencias significativas entre las 3 comunidades de ichneumónidos. Las estructuras comunitarias dependen de la distribución biogeográfica de las especies y del mes de recolecta, mientras que las estructuras poco dependientes, del tipo de vegetación. Tres especies

obtenidas en este estudio representan nuevos registros para México y 5 para el estado de Tamaulipas.

Palabras clave: Análisis de correspondencia; Avispas parasitoides; Neotrópico; Manglar; Provincias biogeográficas

Introduction

The insect order Hymenoptera includes 3 large groups of parasitic wasps: Chalcidoidea and Braconidae, which are more speciose in the tropics (Horstmann et al., 1999; Noyes, 1989; Wharton et al., 1997) and Ichneumonidae, which are mostly found in temperate zones (Jones et al., 2012; Owen & Owen, 1974; Skillen et al., 2000). Although some authors consider ichneumonids to be an exception to the general trend that on a large-scale species richness increases at lower latitudes (Owen & Owen, 1974; Veijalainen et al., 2012), others have questioned this hypothesis because of description and sampling biases, since small koinobiont species might be overlooked or undersampled in the tropics (Quicke, 2012; Santos & Quicke, 2011; Veijalainen et al., 2013). The abundance and distribution of ichneumonids, similar to all organisms, is determined by climate and availability of resources. However, biodiversity studies are not always related to the sources of variation that influence on distribution patterns of species, such as vegetation types, seasonal climatic changes, and biogeographic distribution (Brown & Maurer, 1989; Clark et al., 2011; Menezes et al., 2015; Poulin et al., 2011; Sääksjärvi et al., 2006; Wolda, 1988).

With a global estimate of 100,000 species (Gauld, 2000), of which more than 24,281 have been described (Yu et al., 2015), the Ichneumonidae is one of the most species-rich animal families in the world (Goulet & Huber, 1993; Triplehorn & Johnson, 2005; Veijalainen et al., 2012). Nearly 7,400 ichneumonid species have been defined as Neotropical, and approximately 7,700 species have been described as Nearctic. In Mexico, where the Nearctic and Neotropical regions converge, approximately 1,300 ichneumonid species have been recorded (Ruiz-Cancino, 2015), of which 59%, 29%, and 10% have Neotropical, both Neotropical and Nearctic and Nearctic distributions, respectively, whereas the remaining species show other trends (Ruiz-Cancino et al., 2014). Extensive faunistic studies of Ichneumonidae have been carried out in Tamaulipas, Veracruz, and Yucatan, which are coastal states on the Gulf of Mexico (Ruiz-Cancino, 2015). However, the fauna of Ichneumonidae have been poorly studied in coastal habitats such as mangroves, which are interesting ecosystems for studying parasitic wasps (Myartseva et al., 2014).

In this study, the community structure of Ichneumonidae in a mangrove area in the southern region of Tamaulipas,

Mexico, is described and represents the first study of this family in such habitat. Also, 2 nearby localities with different vegetation types were examined. The study area is characterized as Neotropical, but it is actually on the boundary of the Nearctic region in the northeastern region of the country. Therefore, in this study we evaluated the diversity and species composition of ichneumonid species depending on their biogeographical distribution, type of vegetation, and month of collection.

Materials and methods

Three localities were sampled, which belong to the municipality of Altamira in the state of Tamaulipas, Mexico, at an elevation of 2-15 m asl. These localities correspond to a mangrove area in the Municipality Armenta ($22^{\circ}25'48.30''$ N, $97^{\circ}52'34.86''$ W), a residual low deciduous forest (LDF) in the Congregación Las Prietas ($22^{\circ}31'46.53''$ N, $97^{\circ}56'52.86''$ W), and a low thorny forest (LTF) in the Municipality Aquiles Serdan ($22^{\circ}33'2.87''$ N, $97^{\circ}54'13.11''$ W). The LDF is lush green with dense foliage and a thick herbaceous layer during the rainy season and after the arrival of summer season most trees shed their leaves. The dominant plant genera are *Bursera*, *Phoebe*, *Chloroleucon*, and *Acacia* (Rzedowski, 2006). The LTF is present in drier sites than those where LDF occurs, with predominance of either evergreen or deciduous thorny trees. The dominant plant genera are *Prosopis*, *Esenbeckia*, *Pithecellobium*, and *Acacia* (Rzedowski, 2006). The distance among sites is 12-14 km. The weather in the region is predominantly warm and humid with rain in summer. The monthly mean temperature varies from 16.4 °C in January to 25.2 °C in May. The monthly average rainfall varies from 1.8 mm in February to 206.5 mm in September (Table 1) (Conagua, 2012, 2013).

Wasps were captured from May 2012 to April 2013 with Malaise traps that were fabricated based on the design published by Townes (1972). The height of the alcohol pot was approximately 2 m. The Malaise trap color was black, and measured 1.8 m wide by 1.1 m high with a depth of 60 cm for each side. The plastic pot was half full of 95% ethyl alcohol and the samples were collected every 15 days and conserved in refrigeration until examination. One trap was placed in each site. All traps were placed in open areas, oriented with the collector pot toward the east.

Taxonomic determination was carried out using

Table 1

Monthly temperature and rainfall.

Months	Average monthly temperature (°C)	Monthly rainfall (mm)
May	25.2	14.8
June	24	30.7
July	24.9	115.6
August	24.5	81
September	24.1	206.5
October	21.1	23.6
November	18.7	45.3
December	18.6	2
January	16.4	11.5
February	19.1	1.8
March	17.1	30.3
April	21.2	3

the relevant keys (Brambila, 1997; Dasch, 1979, 1984, 1988, 1992; Gauld, 1988, 1991, 1997, 2000; Gauld et al., 2002; Heinrich, 1977; Kasparyan & Ruíz-Cancino, 2005, 2007, 2008a, 2008b; Khalaim & Ruíz-Cancino, 2012; Reshchikov; 2011; Townes, 1969, 1970a, 1970b, 1971; Townes & Townes, 1978). The specimens that did not match with the morphological features of a described species were regarded as morphospecies. All insects were deposited in the collection of Instituto Tecnológico de Ciudad Victoria, Tamaulipas, Mexico.

Data analysis

Species accumulation curves were generated for each site using the Clench model, which is an adaptation of the Michaelis-Menten equation that is used to describe the kinetics of enzymatic reactions (Jiménez-Valverde & Hortal, 2003). Because the sites are small, all species have the same probability of being caught, and thus we used the negative exponential equation Clench model (Soberón & Llorente, 1993). The accumulation of species as a function of effort was obtained by the Mao Tau estimator, and the adjustment of the Clench equation was performed with the Quasi-Newton simplex algorithm (StatSoft Statistica 8.0). Alpha diversity was determined at each site from the richness and abundance values using the Simpson index of species evenness and the Shannon-Wiener index of diversity (Magurran, 2004). A t-test was carried out to determine statistically significant differences in Simpson Index data. The Berger-Parker dominance index was used to estimate the proportions of the most abundant species (Moreno, 2001). Differences among sites in the collected species and their abundances were determined

by a Permanova multifactorial analysis, which is employed to test the simultaneous response of 1 or more variables to 1 or more factors using permutation methods. This statistic test directly uses the distance matrix to partition the diversity among sources of variation, and is especially suitable for analysis of composition data from ecological studies (Tang et al., 2016). The distance index used was Sorenson-Dice because it assumes imperfect sampling. This analysis was performed with PAST software (version 3.0; Anderson, 2005).

The association of species on type of vegetation, month of collection, and their assigned biogeographical distribution was performed with a multiple correspondence analysis (Abdi & Valentine, 2007; Legendre & Legendre, 2003). The biogeographical distribution of each species was obtained from Yu et al. (2015), and those cataloged as Neotropical-endemic to Mexico (Nt-End) were based on Ruiz-Cancino (2010). The multiple correspondence analysis is a modification of χ^2 used to analyze multiple categorical variables and creates a Cartesian diagram based on the association among more than 2 categorical variables (Legendre & Legendre, 2003). The diagram displays simultaneously the relative position of the categories of variables studied (Gotelli, 2001). The positions of the categories reflect the degree of association among them, nearest categories of different variable represent a high association, while distant categories show lower association. This analysis was performed with the program Statistica version 12 (StatSoft, 2007).

Results

A total of 770 specimens of Ichneumonidae belonging to 19 subfamilies, 66 genera, and 129 species were collected at the 3 localities (Appendix). The overall capture rate for the study was 0.70 individuals per day-trap and the capture rate of species relative to specimens was 16.8 per 100. The highest relative abundance, with 49.6% of all wasps, was observed in the mangrove habitat; the relative abundances in the LTF and the LDF were 25.7% and 24.7%, respectively. The fewest number of specimens for the 3 sites was obtained in January (25 wasps), whereas March was the most abundant month (107 specimens). The months with more specimens were August, July, and March for the mangrove habitat, LDF, and LTF, respectively. Cryptinae was the most abundant subfamily, represented by 39.6% of the collected specimens (Table 2). Cryptinae was common in the 3 sites, whereas Rhyssinae was only collected in LDF. Specimens of Brachycyrtinae, Mesochorinae, and Metopiinae subfamilies were only collected in LTF. The most abundant species were *Anomalon ejuncidum* Say (72 specimens), *Eudeleboea subflava* Davis (42 specimens),

and *Pachysomoides stupidus* Cresson (42 specimens) (Appendix).

The mangrove area had the highest richness with 88 species of which 31 were exclusive for this site. Sixty-four species (17 exclusive) were collected in LDF, whereas 61 species were collected in LTF (18 were exclusive) (Appendix). The most species-rich subfamilies were Cryptinae (45 species), Cremastinae and Ichneumoninae (both with 16 species), and Campopleginae (14 species). The most species-rich genera were *Eiphosoma* with 8 species, and *Lymeon* and *Carinodes*, with 7 species each.

The maximum values of the Clench function curve for each site are presented in Table 3. The coefficient

of determination was greater than 0.9 in all 3 cases, indicating the data were properly matched to the function. This analysis showed that the mangrove site, with 102 estimated species, could potentially be the richest in ichneumonid species of the 3 collected sites. The number of estimated species for the 2 other vegetation types was of 76 in both cases (Table 3).

Comparison of the Ichneumonidae communities at the 3 locations by Permanova showed statistically significant differences ($F = 1.95, p = 0.0004$) among the sites (Table 4). Thus, the 129 species collected in the study area have clearly different spatial distributions.

These community differences at the 3 sites were not

Table 2

Abundance and species richness per subfamily in the 3 localities.

	Low deciduous forest		Mangrove		Low thorny forest		Total	
	Abundance	Richness	Abundance	Richness	Abundance	Richness	Abundance	Richness
Anomaloninae	18	2	49	4	30	4	97	5
Banchinae	8	2	19	2	19	1	46	2
Brachycyrtinae	0	0	2	2	0	0	2	2
Campopleginae	7	3	37	12	7	5	51	14
Cremastinae	8	6	20	8	15	6	43	16
Cryptinae	56	24	176	33	73	23	305	45
Ctenopelmatinae	2	1	2	1	1	1	5	1
Ichneumoninae	27	9	21	8	22	9	70	16
Labeninae	24	4	12	4	5	2	41	5
Lycorininae	1	1	2	1	0	0	3	1
Mesochorinae	0	0	0	0	1	1	1	1
Metopiinae	0	0	0	0	1	1	1	1
Nesomesochorinae	7	1	10	1	8	1	25	1
Ophioninae	9	2	12	2	5	3	26	5
Orthocentrinae	4	2	3	2	9	2	16	2
Pimplinae	11	5	15	6	1	1	27	9
Rhyssinae	3	1	0	0	0	0	3	1
Tersilochinae	0	0	1	1	0	0	1	1
Tryphoninae	5	1	1	1	1	1	7	1

Table 3

Data from Clench model at the 3 sites.

	Low deciduous forest	Mangrove	Low thorny forest
Collected species	64	88	61
Percentage of the collected biota	84.03	62.75	80.46
Estimated species	76.15	101.98	75.81
Coefficient of determination (R)	0.9996	0.99894	0.99895

detected in terms of the proportional abundance of each species. That is, the Shannon-Wiener and Simpson indexes were similar for the 3 data sets (Table 5). The t-test carried out on the Simpson index data did not indicate a significant difference between the mangrove and LDF ($t_{(434)} = -0.924$; $p > 0.05$) or between LDF and LTF sites ($t_{(387)} = 1.245$; $p > 0.05$); the only statistically significant difference was detected by comparing the alpha diversity of the mangrove and LTF ($t_{(445)} = 2.277$; $p < 0.05$). This indicates that the probability of taking 2 individuals of the same species was greater in LTF than in mangrove, and the uncertainty of knowing the identity of a species was greater in mangrove than in LTF. The Berger-Park index, which expresses the number of individuals in proportion to the most abundant species, did not differ between locations (Table 5).

The multiple correspondence analysis (Fig. 1) revealed that biogeographical regions, month of collection, and localities were statistically significant factors ($X^2_{d.f.=24,964} = 149,015$; $p = 0.000$) in the community structure of the Ichneumonidae species. The position of categories of biogeographical regions and identity of species was far from the center of graph, and therefore contributes with most of the observed variation. In contrast, the types of vegetation were closer to the center of graph, which means a lower variation. The month of collection had more influence than sites; for example, species cataloged to Nt-End and Nt-Ne regions were most frequent from February to June, and those species from Ne-Nt-Oc-Pa, Ne-Pa-Nt or Nt-Ne-Oc regions were mostly present on August or October. However, some species did not have correspondence with any month or type of vegetation but only with their recorded biogeographical distribution.

Three species collected in this study represent new records for Mexico: *Pristomerus jugulator* Gauld, 2000, *Physotarsus emarginatus* Zhaurova, 2009, and *Apechoneura mariae* Gauld, 2000. Five species, *Brachycyrtus cosmetus* (Walker, 1956); *Pristomerus sulcatus* (Cameron, 1908); *Temelucha hilux* Gauld, 2000; *Chirotica confederatae* (Ashmead, 1896); and *Enicospilus kleini* Gauld, 1988, are new records for Tamaulipas.

Discussion

This study showed that the community composition of the Ichneumonidae in a mangrove area may significantly differ from nearby areas with different types of vegetation, such as LDF and LTF. In addition, the community structure in this group tend to be determined mostly by the biogeographical distribution of species, followed by the temporal variation throughout the year and less by the type of vegetation. Using multivariate analysis techniques, this study provides a relevant contribution for the Ichneumoid diversity in mangrove areas (Burrows, 2003).

The community of ichneumonids in the mangrove was different in comparison with the other 2 sites. First, the species assemblages of the 3 types of vegetation were significantly different. Second, the abundance and richness were highest in the mangrove area. Third, the local diversity in mangrove was significant higher than LTF, although not different with LDF.

Malaise traps represent a frequently used collecting technique for sampling Ichneumonidae (Fraser et al., 2008; Hall et al., 2015; Pérez-Urbina et al., 2010; Sääksjärvi et al., 2006; Townes, 1972). The overall capture rate for

Table 4

P values from Permanova analysis.

Sites	Low deciduous forest	Mangrove	Low thorny forest
Low deciduous forest	-	0.0168	0.0054
Mangrove	0.0168	-	0.0408
Low thorny forest	0.0054	0.0408	-

Table 5

Abundance, shared species, exclusive species, and diversity indexes at each site.

Sites	Abundance	Richness of species	Shared species	Exclusive species	Shannon-Wiener	Simpson	Berger-Parker
Low deciduous forest	190	64	47 (73.4%)	17 (26.6%)	3.752	0.0355	0.11053
Mangrove	382	88	57 (64.8%)	31 (35.2%)	3.841	0.0372	0.1021
Low thorny forest	198	61	43 (70.5%)	18 (29.5%)	3.621	0.0401	0.096

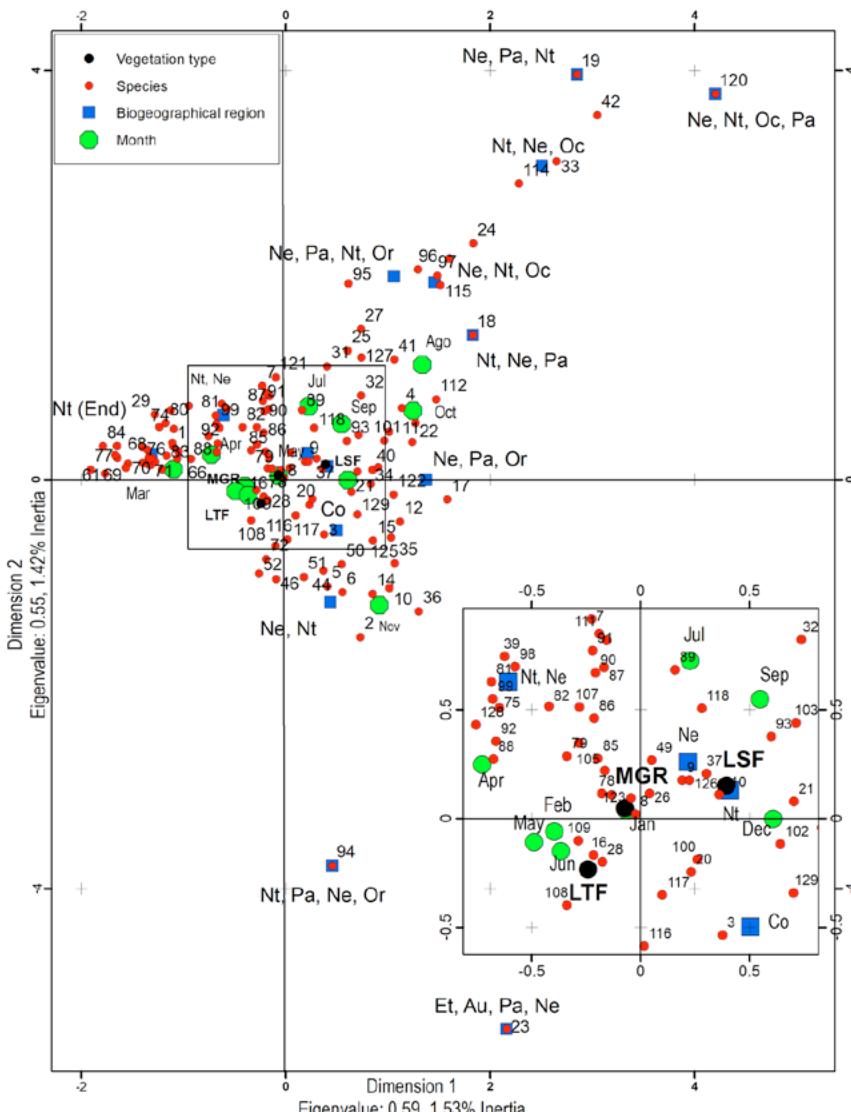


Figure 1. Multivariate correspondence analysis. Ne: Nearctic; Nt: Neotropical; Pa: Palearctic; Or: Oriental; Oc: Oceanic; Co: Cosmopolitan; Et: Ethiopian; Au: Australian; End: Endemic to Mexico; MGR: Mangrove; LDF: Low-Stature Deciduous Forest; LTF: Low-Stature Thorny Forest.

the study was 0.70 individuals per day-trap. This value is in agreement with those reported in other studies, which vary from 2.1 wasps per day-trap (Pérez-Urbina et al., 2010) to low-rate collections of 0.22 individuals per day (Sääksjärvi et al., 2006). However, the capture rate of species relative to specimens was 16.8 per 100. The values found in other studies range from 7 to 17 species per 100 ichneumonids caught (Hall et al., 2015; Pérez-Urbina et al., 2010; Sääksjärvi et al., 2006; Townes, 1971). In comparison, a study with pan traps captured 0.22 species per 100 specimens from a total of 7,500 ichneumonid

wasps collected (Gould et al., 2013). Thus, the values of abundance and richness in the present study using Malaise traps are among those obtained by other studies and allowed the determinations of significant statistical differences.

Mangroves occur in tropical and subtropical latitudes along coastal intertidal areas (Burrows, 2003). In Mexico, they are distributed inside coastal lagoons and deltaic systems (López & Ezcurra, 2002). Mangroves are forests of salt-tolerant woody plants, often comprising 1 species or a mixture of 3 species of these trees (Burrows, 2003). Although their main feature is general floristic simplicity,

mangroves are ecosystems of crucial importance for the life cycles of aquatic fauna (Kathiresan & Bingham, 2001). The legal protection in Mexico is based on their ecological relevance (Moral-Padilla et al., 2012). Entomofaunistic studies of these ecosystems usually assess plant-insect relationships through herbivory (Burrows, 2003; Feller & Chamberlain, 2007; Feller & Mathis, 1997; Murphy & Lugo, 1986). However, there are few studies in other trophic guilds. The only faunistic study of parasitoid and predatory insects in mangroves was carried out by Veenakumari et al. (1997) on 2 Indian islands. These authors suggested that these kinds of forests cannot sustain a large population of insects, but they are influenced by their adjacent areas (Berjak et al., 1977; Kathiresan & Bingham, 2001; Veenakumari et al., 1997). Thus, the richness of Ichneumonidae collected in the mangrove area in this study could result from the mutual influences of the mangrove and its surroundings. Rao et al. (1998) had already mentioned that mangrove insect visitors could function as links between these aquatic ecosystems and neighboring areas.

In this study, the community structure of Ichneumonidae was strongly determined by the biogeographical distribution where species were reported. The relative abundances of species showed strongest correspondence with the biogeographical categories compared to collecting month and type of vegetation. Neotropical (Nt, Nt-Ne, Nt-End) species, which were the most abundant, were collected throughout all months of the year, although their frequency was lower in August, October, and November. The frequency of Nt-End and Nt-Ne species was associated with the months from February to June, when the temperature

ranged from 19.1-24 °C and the rainfall from 1.8 mm to 30.7 mm. In comparison, the most-rainy months were July (311 mm) and September (314 mm), but their association with the relative abundance of the species was lower, since the associated species tend to be cosmopolites. With respect to types of vegetation, the correspondence was lower than the collecting month. These data indicate that the zone is markedly Neotropical with a good established community of ichneumonids, despite fluctuations of temperature and precipitation through the year.

In conclusion, the community structure of the ichneumonid wasps in the subtropical zones near to Nearctic region strongly depends on the biogeographical distribution of the species in association with climatic variation throughout the year. However, the mangrove and their surroundings appear to be favorable for this group of insects, which is a relevant issue given the limited information of parasitic Hymenoptera in this ecosystem.

Finally, with the new records for Mexico and Tamaulipas presented here, knowledge about the geographical distribution of some species has been extended. Based on this, 1,306 species of Ichneumonidae fauna are now registered for Mexico (Ruiz-Cancino, 2015) and 417 species are reported for the state of Tamaulipas (Ruiz-Cancino, 2010).

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Appendix. List of species of Ichneumonidae collected in this study. Nt: Neotropical, Ne: Nearctic, Pa: Palaearctic, Or: Oriental, Oc: Oceanic, Co: Cosmopolitan, Et: Ethiopic, Au: Australian, End: Endemic to Mexico. LDF: low-stature deciduous forest, MGR: mangrove, LTF: low-stature thorny forest. *New records to Tamaulipas, **New records to Mexico.

	Subfamily Species/Morphospecies	Biogeography	LDF	MGR	LTF	♀/♂	Total
I.	Anomaloninae						
1	<i>Anomalon constrictum</i> Dasch, 1984	Nt, Ne	0	0	5	5/0	5
2	<i>Anomalon ejuncidum</i> Say, 1835	Ne, Nt	16	38	18	72/0	72
3	<i>Anomalon fuscipes</i> (Cameron, 1886)	Nt	0	1	1	2/0	2
4	<i>Corsoncus minori</i> Gauld y Bradshaw, 1997	Nt	0	1	0	1/0	1
5	<i>Ophiopterus cincticornis</i> (Cresson, 1865)	Ne, Nt	2	9	6	16/1	17
II.	Banchinae						
6	<i>Diradops hyphantrinae</i> Kasparyan y Pinson, 2007	Nt (End)	2	2	0	4/0	4
7	<i>Meniscomorpha subflava</i> (Davis, 1898)	Ne, Nt	6	17	19	30/12	42
III.	Brachycyrtinae						

Appendix. Continued...

	Subfamily Species/Morphospecies	Biogeography	LDF	MGR	LTF	♀/♂	Total
8	<i>Brachycyrtus cosmetus</i> (Walkley, 1956)*	Nt	0	1	0	1/0	1
9	<i>Brachycyrtus pretiosus</i> Cushman, 1936	Nt	0	1	0	1/0	1
IV.	Campopleginae						
10	<i>Campoletis</i> sp. Förster, 1869	Co	0	5	0	5/0	5
11	<i>Casinaria</i> sp. 1 Holmgren, 1859	Co	0	2	1	2/1	3
12	<i>Casinaria</i> sp. 2 Holmgren, 1859	Co	0	2	1	1/2	3
13	<i>Casinaria</i> sp. 3 Holmgren, 1859	Co	0	0	2	2/0	2
14	<i>Diadegma</i> sp. 1 Förster, 1869	Co	0	2	0	2/0	2
15	<i>Diadegma</i> sp. 2 Förster, 1869	Co	0	2	0	1/1	2
16	<i>Dusona</i> sp. Cameron, 1901	Co	0	1	0	1/0	1
17	<i>Echthronomas</i> sp. Förster, 1869	Ne, Pa, Or	1	5	0	6/0	6
18	<i>Microcharops</i> sp. Roman, 1910	Nt, Ne, Pa	4	11	0	13/2	15
19	<i>Phobocampe</i> sp. Förster, 1869	Ne, Pa, Nt	2	0	2	4/0	4
20	<i>Venturia</i> sp. 1 Schrottky, 1902	Co	0	4	1	5/0	5
21	<i>Venturia</i> sp. 2 Schrottky, 1902	Co	0	1	0	1/0	1
22	<i>Venturia</i> sp. 3 Schrottky, 1902	Co	0	1	0	1/0	1
23	<i>Xanthocampoplex</i> sp. Morley, 1913	Et, Au, Pa, Ne	0	1	0	1/0	1
V.	Cremastinae						
24	<i>Eiphosoma</i> sp. 1 Cresson, 1865	Nt, Ne, Oc	0	0	1	1/0	1
25	<i>Eiphosoma</i> sp. 2 Cresson, 1865	Nt, Ne, Oc	0	0	1	1/0	1
26	<i>Eiphosoma batatae</i> Cushman, 1931	Nt	0	2	0	2/0	2
27	<i>Eiphosoma dentator</i> (Fabricius, 1804)	Nt, Ne, Oc	0	1	1	2/0	2
28	<i>Eiphosoma macrum</i> (Enderlein, 1921)	Nt	0	0	3	2/1	3
29	<i>Eiphosoma mexicanum</i> Cresson, 1874	Nt, Ne	0	1	0	0/1	1
30	<i>Eiphosoma nigrolineatum</i> (Brullé, 1846)	Nt, Ne	1	0	0	0/1	1
31	<i>Eiphosoma vitticolle</i> Cresson, 1865	Nt, Ne	0	2	0	1/1	2
32	<i>Pristomerus jugulator</i> Gauld, 2000**	Nt	1	0	0	0/1	1
33	<i>Pristomerus spinator</i> (Fabricius, 1804)	Ne, Nt, Oc	0	1	0	1/0	1
34	<i>Pristomerus sulcatus</i> (Cameron, 1908)*	Nt	0	8	3	10/1	11
35	<i>Temelucha hilux</i> Gauld, 2000*	Nt	1	1	0	2/0	2
36	<i>Temelucha</i> sp. Förster, 1869	Co	1	0	0	1/0	1
37	<i>Xiphosomella dubia</i> (Brues, 1911)	Ne	1	4	0	2/3	5
38	<i>Xiphosomella</i> sp. 1 Szépligeti, 1905	Nt, Ne	0	0	6	5/1	6
39	<i>Xiphosomella</i> sp. 2 Szépligeti, 1905	Nt, Ne	3	0	0	2/1	3
VI.	Cryptinae						
40	<i>Acerastes accolens</i> (Cresson, 1874)	Nt (End)	1	5	0	2/4	6
41	<i>Acerastes faciator</i> Kasparyan et Ruíz, 2008	Nt (End)	2	0	2	4/0	4
42	<i>Acerastes myartsevae</i> Kasparyan et Ruíz, 2008	Nt (End)	0	0	1	1/0	1
43	<i>Acerastes pertinax</i> (Cresson, 1872)	Nt (End)	2	2	6	3/7	10
44	<i>Acerastes scabrosus</i> Kasparyan et Ruíz, 2008	Nt (End)	2	0	4	3/3	6
45	<i>Acerastes tinctor</i> Kasparyan et Ruíz, 2008	Nt (End)	0	0	7	7/0	7
46	<i>Agonocryptus chichimecus</i> (Cresson, 1874)	Nea, Neo	5	17	16	18/20	38

Appendix. Continued...

	Subfamily Species/Morphospecies	Biogeography	LDF	MGR	LTF	♀/♂	Total
47	<i>Baryceros zapotecus</i> (Cresson, 1874)	Nt (End)	0	2	0	2/0	2
48	<i>Basileucus suspiciosus</i> Kasparyan et Ruíz, 2008	Nt (End)	0	1	0	1/0	1
49	<i>Cestrus arcuatorius</i> Kasparyan et Ruíz, 2005	Nt (End)	10	3	0	4/9	13
50	<i>Chirotica confederatae</i> (Ashmead, 1896)*	Ne	0	1	1	2/0	2
51	<i>Cryptanura brachygaster</i> (Cameron, 1885)	Nt (End)	0	5	1	3/3	6
52	<i>Cryptanura lunai</i> Kasparyan y Ruíz, 2006	Nt (End)	0	0	1	0/1	1
53	<i>Diapetimorpha acadia</i> Cushman, 1929	Ne, Nt	1	0	0	1/0	1
54	<i>Diapetimorpha communis</i> (Cresson, 1874)	Ne, Nt	2	1	0	1/2	3
55	<i>Diapetimorpha macula</i> (Cameron, 1886)	Ne, Nt	5	1	4	10/0	10
56	<i>Diapetimorpha monilis</i> (Cresson, 1874)	Nt	0	1	0	1/0	1
57	<i>Diapetimorpha pedator</i> Kasparyan et Ruíz, 2005	Nt (End)	1	0	1	0/2	2
58	<i>Diapetimorpha scitula</i> (Cresson, 1874)	Nt (End)	3	4	5	8/4	12
59	<i>Digonocryptus thoracicus</i> Kasparyan et Ruíz, 2005	Nt	4	11	0	10/5	15
60	<i>Digonocryptus variegatus</i> (Szépligeti, 1916)	Nt	0	1	0	1/0	1
61	<i>Epicnemion lineator</i> Kasparyan et Ruíz, 2008	Nt (End)	0	1	0	1/0	1
62	<i>Gambrus bituminosus</i> (Cushman, 1924)**	Ne	0	1	0	1/0	1
63	<i>Gambrus ultimus</i> (Cresson, 1864)	Ne, Nt, Oc	1	0	0	1/0	1
64	<i>Joppidium brochum</i> Townes, 1962	Ne, Nt	4	0	0	1/3	4
65	<i>Latosculum ruizi</i> Kasparyan, 2004	Nt (End)	0	3	0	1/2	3
66	<i>Lymeon acceptus</i> (Cresson, 1874)	Nt (End)	0	1	1	1/1	2
67	<i>Lymeon mandibularis</i> Kasparyan y Ruíz, 2004	Nt (End)	1	0	0	0/1	1
68	<i>Lymeon minutus</i> Kasparyan y Ruíz, 2008	Nt (End)	1	10	4	14/1	15
69	<i>Lymeon moratus</i> (Cresson, 1874)	Nt (End)	3	22	5	15/15	30
70	<i>Lymeon rufotibialis</i> Kasparyan y Ruíz, 2004	Nt (End)	0	1	3	4/0	4
71	<i>Lymeon tantillus</i> (Cresson, 1874)	Nt (End)	2	5	1	5/3	8
72	<i>Lymeon tricoloripes</i> Kasparyan y Ruíz, 2004	Nt (End)	1	1	1	2/1	3
73	<i>Mallochia</i> sp. Viereck, 1912	Ne, Nt	1	0	0	0/1	1
74	<i>Messatoporus compressicornis</i> Cushman, 1929	Ne, Nt	1	12	1	9/5	14
75	<i>Messatoporus discoidalis</i> (Cresson, 1872)	Ne, Nt	1	0	0	0/1	1
76	<i>Messatoporus mesonotator</i> Kasparyan y Ruíz, 2005	Nt (End)	0	2	0	2/0	2
77	<i>Messatoporus propodeator</i> Kasparyan y Ruíz, 2005	Nt (End)	0	1	0	0/1	1
78	<i>Pachysomoides stupidus</i> (Cresson, 1874)	Nt, Ne	0	39	3	42/0	42
79	<i>Polycyrtidea carlosi</i> Kasparyan et Ruíz, 2008	Nt (End)	1	2	0	1/2	3
80	<i>Polycyrtidea limitis</i> Cushman, 1929	Nt, Ne	0	2	0	2/0	2
81	<i>Polycyrtus melanoleucus</i> (Brullé, 1846)	Nt	1	4	1	3/3	6
82	<i>Polycyrtus semialbus</i> (Cresson, 1865)	Nt	0	11	0	8/3	11
83	<i>Polycyrtus</i> sp. Spinola, 1840	Nt, Ne	0	1	3	4/0	4
84	<i>Xenarthron pectoralis</i> Kasparyan et Ruíz, 2005	Nt (End)	0	2	1	3/0	3
VII.	Ctenopelmatinae						
85	<i>Physotarsus emarginatus</i> Zhaurova, 2009**	Ne	2	2	1	5/0	5
VIII.	Ichneumoninae						
86	<i>Carinodes</i> sp. 1 Hancock, 1926	Nt, Ne	8	2	9	18/1	19

Appendix. Continued...

	Subfamily Species/Morphospecies	Biogeography	LDF	MGR	LTF	♀/♂	Total
87	<i>Carinodes</i> sp. 2 Hancock, 1926	Nt, Ne	3	5	0	7/1	8
88	<i>Carinodes</i> sp. 3 Hancock, 1926	Nt, Ne	2	0	5	0/7	7
89	<i>Carinodes</i> sp. 4 Hancock, 1926	Nt, Ne	3	0	0	3/0	3
90	<i>Carinodes</i> sp. 5 Hancock, 1926	Nt, Ne	2	0	0	0/2	2
91	<i>Carinodes</i> sp. 6 Hancock, 1926	Nt, Ne	0	0	1	1/0	1
92	<i>Carinodes</i> sp. 7 Hancock, 1926	Nt, Ne	0	0	1	0/1	1
93	<i>Lobaegis arista</i> (Cresson, 1868)	Nt (End)	2	3	0	3/2	5
94	<i>Oedicephalus</i> sp. Cresson, 1868	Nt	1	1	1	2/1	3
95	<i>Phaeogenes</i> sp. Wesmael, 1845	Ne, Pa, Or	0	3	0	3/0	3
96	<i>Projoppa</i> sp. Townes, 1966	Nt	4	4	1	7/2	9
97	<i>Tricholabus</i> sp. 1 Thomson, 1894	Nt, Pa, Ne, Or	0	0	2	2/0	2
98	<i>Tricholabus</i> sp. 2 Thomson, 1894	Ne, Pa, Nt, Or	0	2	0	0/2	2
99	<i>Tricholabus</i> sp. 3 Thomson, 1894	Ne, Pa, Nt, Or	0	1	1	0/2	2
100	<i>Tricholabus</i> sp. 4 Thomson, 1894	Ne, Pa, Nt, Or	0	0	1	1/0	1
101	<i>Trogomorpha arrogans</i> (Cresson, 1874)	Nt, Ne	2	0	0	1/1	2
IX.	Labeninae						
102	<i>Apechoneura mariae</i> Gauld, 2000**	Nt	1	1	0	2/0	2
103	<i>Grotea fulva</i> Cameron, 1886	Nt	1	3	0	4/0	4
104	<i>Labena eremica</i> Gauld, 2000	Nt	20	6	3	10/19	29
105	<i>Labena schausi</i> Cushman, 1922	Nt	0	2	0	2/0	2
106	<i>Labena tarsata</i> Gauld, 2000	Nt	2	0	2	2/2	4
X.	Lycorinae						
107	<i>Lycorina apicalis</i> Cresson, 1874	Nt, Ne	1	2	0	2/1	3
XI.	Mesochorinae						
108	<i>Mesochorus</i> sp. Gravenhorst, 1829	Co	0	0	1	1/0	1
XII.	Metopiinae						
109	<i>Metopius errantia</i> Davis, 1897	Ne	0	0	1	0/1	1
XIII.	Nesomesochorinae						
110	<i>Nonnus</i> sp. Cresson, 1874	Nt	7	10	8	9/16	25
XIV.	Ophioninae						
111	<i>Enicospilus glabratu</i> s (Say, 1835)	Nt, Ne	0	4	0	4/0	4
112	<i>Enicospilus kleini</i> Gauld, 1988*	Nt	1	0	0	1/0	1
113	<i>Enicospilus masoni</i> Gauld, 1988*	Nt	0	0	1	1/0	1
114	<i>Enicospilus purgatus</i> (Say, 1835)	Ne, Nt, Oc	0	0	3	3/0	3
115	<i>Enicospilus trilineatus</i> (Brullé, 1846)	Nt, Ne, Oc	8	8	1	14/3	17
XV.	Orthocentrinae						
116	<i>Megastylus</i> sp. Schiødte, 1838	Co	1	2	8	10/1	11
117	<i>Orthocentrus</i> sp. Gravenhorst, 1829	Co	3	1	1	5/0	5
XVI.	Pimplinae						
118	<i>Clistopyga calixtoi</i> Gauld, 1991	Nt	0	0	1	1/0	1
119	<i>Clydonium quintanillai</i> Gauld, 1991	Nt	0	2	0	2/0	2

Appendix. Continued...

	Subfamily Species/Morphospecies	Biogeography	LDF	MGR	LTF	♀/♂	Total
120	<i>Neotheronia melloso</i> (Cresson, 1874)	Nt	4	0	0	2/2	4
121	<i>Neotheronia tacubaya</i> (Cresson, 1874)	Nt	0	2	0	0/2	2
122	<i>Pimpla punicipes</i> Cresson, 1874	Nt, Ne, Oc	3	4	0	0/7	7
123	<i>Pimpla sanguinipes</i> Cresson, 1872	Ne, Nt	2	4	0	5/1	6
124	<i>Pimpla sumichrasti</i> Cresson, 1874	Nt	1	2	0	2/1	3
125	<i>Scambus</i> sp. Hartig, 1838	Ne, Nt, Oc, Pa	1	0	0	0/1	1
126	<i>Zatypota alborhombarta</i> (Davis, 1895)	Nt, Ne	0	1	0	1/0	1
XVII.	Rhyssinae						
127	<i>Epirhyssa mexicana</i> Cresson, 1874	Nt, Ne	3	0	0	0/3	3
XVIII.	Tersilochinae						
128	<i>Allophrys divisoricata</i> Horstmann, 2010	Nt, Ne	0	1	0	1/0	1
XIX.	Tryphoninae						
129	<i>Netelia</i> sp. Gray, 1860	Co	5	1	1	7/0	7
		Total	190	382	198	555/215	770

References

- Abdi, H., & Valentine, D. (2007). *Multiple correspondence analysis*. In N. J. Salkinds (Eds.), *Encyclopedia of Measurement and Statistics* (pp. 651–657). Thousand Oaks, CA: Sage Publications.
- Anderson, M. J. (2005). *Permanova: A FORTRAN computer program for permutational multivariate analysis of variance*. New Zealand: University of Auckland.
- Berjak, P., Campbell, G. K., Hucket, B. I., & Pammenter, N. W. (1977). *In the mangroves of Southern Africa*. Durban, South Africa: Wildlife Society of Southern Africa.
- Brambila, J. (1997). *Gambrus wileyi* (Hymenoptera: Ichneumonidae) a new Cryptinae wasp from Florida. *Insecta Mundi*, 11, 247–254.
- Brown, J. H., & Maurer, B. A. (1989). Macroecology: the division of food and space among species on continents. *Science*, 243, 1145–1150.
- Burrows, D. W. (2003). *The role of insect leaf herbivory on the mangroves Avicennia marina and Rhizophora stylosa (Thesis)*. James Cook University, Australia.
- Clark, A. T., Rykken, J. J., & Farrell, B. D. (2011). The effects of biogeography on ant diversity and activity on the Boston Harbor Islands, Massachusetts, U.S.A. *Plos One*, 6, e28045.
- Conagua (Comisión Nacional del Agua). (2012). *Banco de información hidroclimatológica*. Estación Altamira, Tamaulipas. Tampico: Gerencia Regional Golfo Norte.
- Conagua (Comisión Nacional del Agua). (2013). *Banco de información hidroclimatológica*. Estación Altamira, Tamaulipas. Tampico: Gerencia Regional Golfo Norte.
- Dasch, C. E. (1979). Ichneumon-flies of America North of Mexico: 8. Subfamily Cremastinae. *Memoirs of the American Entomological Institute*, 29, 1–702.
- Dasch, C. E. (1984). Ichneumon-flies of America North of Mexico: 9. Subfamilies Thriiniiae and Anomaloninae. *Memoirs of the American Entomological Institute*, 29, 1–610.
- Dasch, C. E. (1988). Ichneumon-flies of America North of Mexico: 10. Subfamily Banchinae, tribe Glyptinae. *Memoirs of the American Entomological Institute*, 29, 1–644.
- Dasch, C. E. (1992). Ichneumon-flies of America North of Mexico: 12. Subfamilies Microleptinae, Helictinae, Cylloceriinae and Oxytorinae. *Memoirs of the American Entomological Institute*, 29, 1–470.
- Feller, I. C., & Chamberlain, A. (2007). Herbivore responses to nutrient enrichment and landscape heterogeneity in a mangrove ecosystem. *Oecologia*, 153, 607–616.
- Feller, I. C., & Mathis, W. N. (1997). Primary herbivory by wood-boring insects along an architectural gradient of *Rhizophora* mangle. *Biotropica*, 29, 440–451.
- Fraser, S. E. M., Dytham, C., & Mayhew, P. J. (2008). The effectiveness and optimal use of Malaise traps for monitoring parasitoid wasps. *Insect Conservation and Diversity*, 1, 22–31.
- Gauld, I. D. (1988). A survey of Ophioninae (Hymenoptera: Ichneumonidae) of tropical Mesoamerica with special reference to the fauna of Costa Rica. *Bulletin of the British Museum (Natural History)*. London, 57, 1–309.
- Gauld, I. D. (1991). The Ichneumonidae of Costa Rica, 1. *Memoirs of the American Entomological Institute*, 47, 1–589.
- Gauld, I. D. (1997). The Ichneumonidae of Costa Rica, 2. *Memoirs of the American Entomological Institute*, 47, 1–589.
- Gauld, I. D. (2000). The Ichneumonidae of Costa Rica, 3. *Memoirs of the American Entomological Institute*, 47, 1–589.

- of the American Entomological Institute*, 63, 1–453.
- Gauld, I., Godoy, C., Sithole, R., & Ugalde-Gómez, J. (2002). The Ichneumonidae of Costa Rica, 4. *Memoirs of the American Entomological Institute*, 66, 1–768.
- Gould, R. K., Pejchar, L., Bothwell, S. G., Brosi, B., Wolny, S., Mendenhall, C. D. et al. (2013). Forest restoration and parasitoid wasp communities in montane Hawai'i. *Plos One*, 8, e59356.
- Goulet, H., & Huber, J. T. (1993). *Hymenoptera of the world: an identification guide to families*. Ottawa: Centre for Land and Biological Resources Research.
- Gotelli, N. J. (2001). Research frontiers in null model analysis. *Global Ecology and Biogeography*, 10, 337–343.
- Hall, C. R., Burwell, C. J., Nakamura, A., & Kitching, R. L. (2015). Altitudinal variation of parasitic Hymenoptera assemblages in Australian subtropical rainforest. *Austral Entomology*, 54, 246–258.
- Heinrich, G. H. (1977). Ichneumoninae of Florida and neighboring states (Hymenoptera: Ichneumonidae, subfamily Ichneumoninae). Arthropods of Florida and neighboring land areas. *Florida Department of Agriculture and Consumer Services*, 9, 1–350.
- Horstmann, K., Floren, A., & Linsenmair, K. E. (1999). High species richness of Ichneumonidae (Hymenoptera) from the canopy of a Malaysian rain forest. *Ecotropica*, 5, 1–12.
- Jiménez-Valverde, A., & Hortal, J. (2003). Las curvas de acumulación de especies y la necesidad de evaluar la calidad de los inventarios biológicos. *Revista Ibérica de Aracnología*, 8, 151–161.
- Jones, O. R., Purvis, A., & Quicke, D. L. J. (2012). Latitudinal gradients in taxonomic overdescription rate affect macroecological inferences using species list data. *Ecography*, 35, 333–340.
- Khalaim, A. I., & Ruíz-Cancino, E. (2012). Mexican species of *Exetastes* (Hymenoptera: Ichneumonidae: Banchinae). *Revista Mexicana de la Biodiversidad*, 83, 370–379.
- Kasparyan, D. R., & Ruíz-Cancino, E. (2005). *Avispas parasíticas de plagas y otros insectos, Cryptini de México (Hymenoptera: Ichneumonidae: Cryptinae) Parte I*. Cd. Victoria, Tamaulipas: Universidad Autónoma de Tamaulipas.
- Kasparyan, D. R., & Ruíz-Cancino, E. (2007). Review of Mexican species of *Diradops* with a key and description of a new species (Hymenoptera: Ichneumonidae: Banchinae). *Zoosystematic Rossica*, 16, 263–269.
- Kasparyan, D. R., & Ruíz-Cancino, E. (2008a). *Cryptini de México (Hymenoptera: Ichneumonidae: Cryptinae) Parte II. Serie: Avispas parasíticas de plagas y otros insectos*. Cd. Victoria, Tamaulipas: Universidad Autónoma de Tamaulipas.
- Kasparyan, D. R., & Ruíz-Cancino, E. (2008b). A new species of *Lymeon* from Mexico (Hymenoptera: Ichneumonidae: Cryptini). *Zoosystematic Rossica*, 17, 93–94.
- Kathiresan, K., & Bingham, B. L. (2001). Biology of mangroves and mangrove ecosystems. *Advances in Marine Biology*, 40, 81–251.
- Legendre, P., & Legendre, L. (2003). *Numerical ecology*. Amsterdam: Elsevier.
- López, P. J., & Ezcurra, D. E. (2002). *Los manglares de México: una revisión*. In R. D. Sotelos (Eds.), *Madera y Bosques* (pp. 427–451). Xalapa, Veracruz: Instituto de Ecología, A.C.
- Magurran, A. E. (2004). *Measuring biological diversity*. London: Blackwell Science Ltd.
- Menezes, R. S. T., Brady, S. G., Carvalho, A. F., del Lama, M. A., & Costa, M. A. (2015). Molecular phylogeny and historical biogeography of the neotropical swarm-founding social wasp genus *Synoeca* (Hymenoptera: Vespidae). *Plos One*, 10, e0119151.
- Moral-Padilla, L., Martínez-Puebla, J. A., & Elizalde-Ceballos, S. (2012). La problemática de los manglares, los riesgos naturales en las zonas costeras del sur de Tamaulipas y sus alternativas jurídico administrativas. *Global Conference on Business and Finance Proceedings*, 7, 1408–1418.
- Moreno, C. E. (2001). *Métodos para medir la biodiversidad, Vol. I. M&T-Manuales y Tesis SEA*. Zaragoza, España.
- Murphy, P. G., & Lugo, A. E. (1986). Ecology of tropical dry forest. *Annual Review of Ecology and Systematics*, 17, 67–88.
- Myartseva, S. N., Ruíz-Cancino, E., & Coronado-Blanco, J. M. (2014). Diversidad del género *Encarsia* Förster (Hymenoptera: Chalcidoidea: Aphelinidae) en Tamaulipas, México. *Memorias del Encuentro de la Academia Tamaulipecana de Investigación Científica y Tecnológica, A.C. Tampico, Tamaulipas*, 22–23 de mayo.
- Noyes, J. S. (1989). The diversity of Hymenoptera in the tropics with special reference to Parasitica in Sulawesi. *Ecological Entomology*, 14, 197–207.
- Owen, D. F., & Owen, J. (1974). Species diversity in temperate and tropical Ichneumonidae. *Nature*, 249, 583–584.
- Pérez-Urbina, B. A., Correa-Sandoval, A., Ruíz-Cancino, E., Kasparyan, D. R., Coronado-Blanco, J. M., & Horta-Vega, J. V. (2010). Diversidad de Ichneumonidae (Hymenoptera) en el Cañón del Novillo, Victoria, Tamaulipas, México. *Entomotropica*, 25, 83–97.
- Poulin, R., Krasnov, B. R., Mouillot, D., & Thieltges, D. W. (2011). The comparative ecology and biogeography of parasites. *Philosophical Transactions of the Royal Society B*, 366, 2379–2390.
- Quique, D. L. J. (2012). We know too little about parasitoid wasps distributions to draw any conclusions about latitudinal trends in species richness, body size and biology. *Plos One*, 7, e32101.
- Rao, T. A., Molur, S., & Walker, S. (1998). *Report of the workshop “Conservation assessment and management plan for mangroves of India” (BCPP-Endangered Species Project)*. Coimbatore, India: Zoo Outreach Organization, Conservation Breeding Specialist Group.
- Reshchikov, A. V. (2011). Mexican species of *Lathrolestes* Förster (Hymenoptera: Ichneumonidae: Ctenopelmatinae), with descriptions of four new species and an identification key. *Tijdschrift voor Entomologie*, 154, 15–24.
- Ruiz-Cancino, E. (2010). *Ichneumonidae (Hymenoptera) del Estado de Tamaulipas, México*. Ciudad Victoria, Tamaulipas: Universidad Autónoma de Tamaulipas.
- Ruiz-Cancino, E. (2015). La familia Ichneumonidae

- (Hymenoptera) en México. *Entomología Mexicana*, 2, 1–13.
- Ruiz-Cancino, E., Kasparyan, D. R., González-Moreno, A., Khalaim, A. I., & Coronado-Blanco, J. M. (2014). Biodiversidad de Ichneumonidae (Hymenoptera) en México. *Revista Mexicana de Biodiversidad (Suplem.)*, 85, S385–S391.
- Rzedowski, J. (2006). *Vegetación de México*. México D.F.: Comisión Nacional para el Conocimiento y Uso de la Biodiversidad. Available: http://www.biodiversidad.gob.mx/publicaciones/librosDig/pdf/VegetacionMx_Cont.PDF
- Sääksjärvi, I. E., Ruokolainen, K., Tuomisto, H., Haataja, S., Fine, P. V. A., Cárdenas, G. et al. (2006). Comparing composition and diversity of parasitoid wasps and plants in an Amazonian rainforest mosaic. *Journal of Tropical Ecology*, 22, 167–176.
- Santos, A. M. C., & Quicke, D. L. J. (2011). Large-scale diversity patterns of parasitoid insects. *Entomological Science*, 14, 371–382.
- Skillen, E. L., Pickering, J., & Sharkey, M. J. (2000). Species richness of the Campopleginae and Ichneumoninae (Hymenoptera: Ichneumonidae) along a latitudinal gradient in Eastern North American old-growth forests. *Environmental Entomology*, 29, 460–466.
- Soberón, J., & Llorente, J. (1993). The use of species accumulation functions for the prediction of species richness. *Conservation Biology*, 7, 480–488.
- StatSoft, Inc. (2007). STATISTICA (data analysis software system), version 8.0. www.statsoft.com
- Tang, Z. Z., Chen, G., & Alekseyenko, A. V. (2016). Permanova-S: association test for microbial community composition that accommodates confounders and multiples distances. *Bioinformatics*, 32, 2618–2625.
- Townes, H. (1969). The genera of Ichneumonidae, part 1. *Memoirs of the American Entomological Institute*, 11, 1–300.
- Townes, H. (1970a). The genera of Ichneumonidae, part 2. *Memoirs of the American Entomological Institute*, 12, 1–537.
- Townes, H. (1970b). The genera of Ichneumonidae, part 3. *Memoirs of the American Entomological Institute*, 13, 1–307.
- Townes, H. (1971). The genera of Ichneumonidae, part 4. *Memoirs of the American Entomological Institute*, 17, 372 pp.
- Townes, H. (1972). A light-weight Malaise trap. *Entomological News*, 83, 239–247.
- Townes, H., & Townes, M. (1978). Ichneumon-flies of America North of Mexico, 7. Subfamily Banchinae, tribes Lissonotini and Banchini. *Memoirs of the American Entomological Institute*, 26, 1–614.
- Triplehorn, C. A., & Johnson, N. F. (2005). *Borror and DeLong's introduction to the study of insects*. Belmont, CA.: Thomson-Brooks/Cole.
- Veenakumari, K., Mohanraj, P., & Bandyopadhyay, A. K. (1997). Insect herbivores and their natural enemies in the mangals of the Andaman and Nicobar islands. *Journal of Natural History*, 31, 1105–1126.
- Veijalainen, A., Wahlberg, N., Broad, G. R., Erwin, T. L., Longino, J. T., & Sääksjärvi, I. E. (2012). Unprecedented ichneumonid parasitoid wasp diversity in tropical forests. *Proceedings of the Royal Society B*, 279, 4694–4698.
- Veijalainen, A., Sääksjärvi, I. E., Erwin, T. L., Gómez, I. C., & Longino, J. T. (2013). Subfamily composition of Ichneumonidae (Hymenoptera) from western Amazonia: insights into diversity of tropical parasitoid wasps. *Insect Conservation and Diversity*, 6, 28–37.
- Wharton, R. A., Marsh, P. M., & Sharkey, M. J. (1997). *Manual of the New World genera of the family Braconidae (Hymenoptera)*. Washington, DC: The International Society of Hymenopterists.
- Wolda, H. (1988). Insect seasonality: Why?. *Annual Review of Ecology and Systematics*, 19, 1–18.
- Yu, D. S., van-Achterberg, K., & Horstmann, K. (2015). *World Ichneumonoidea 2011. Taxonomy, biology, morphology and distribution. Taxapad CD/DVD*. Vancouver, Canada. Retrieved on March 9, 2018 from: <http://www.taxapad.com>